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One column to rule them all? (short title)

Current and Future Chromatographic Columns: is one column enough to rule them all? (full title)

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Abstract:

In this contribution, the current state-of-the-art and recent developments in column technology are discussed. The vast majority of separations in liquid chromatography still uses the typical packed particle bed format, most commonly with fully or superficially porous particles in particle sizes as low as 1.3 μm. As an alternative, monolithic columns have been the topic of a large number of studies, but are currently only used in some niche applications. Research into perfectly ordered microfabricated columns has shown tremendous possibilities for these highly performant columns for use in nano-LC, but their development is still ongoing. The possibilities that the emerging technology of 3D printing offers would make it theoretically possible to develop any imaginable structure with high precision, but is currently limited by the available spatial resolution and required printing time, narrowing the size and application range. A critical review of all these technologies is provided and demonstrates that the further development of chromatographic columns will be of paramount importance in the years to come.

30 1. Introduction

31 The improvements in instrument and column performance in liquid chromatography (LC) over
32 the last 15-20 years have resulted in an almost 10-fold reduction in analysis time and 3-fold
33 increase in separation efficiency. Nevertheless, the complexity of the samples emerging in life
34 sciences (proteomics, metabolomics, lipidomics...), containing 10,000 or more analytes in a wide
35 range of concentrations and physico-chemical properties (size, polarity, ionization state...) is so
36 vast that it is impossible to even dream of ever achieving full resolution using unidimensional
37 chromatography with the present state-of-the-art in instrumentation and columns. The rise of
38 two-dimensional separation techniques, in combination with modern MS/MS systems, vastly
39 increases the overall resolving power that can be achieved [1-4]. Nevertheless, even 2D
40 separations are ultimately limited by the efficiency of the chromatographic column in the
41 individual dimensions. In addition to the need for enhanced resolution, a further increase in
42 separation speed for the 2nd dimension column is in high demand as well, as this would allow to
43 increase the sampling rate in the 1st dimension, better preserving its resolution.

44 The further development of faster and more efficient LC chromatographic columns thus remains
45 of paramount importance in the years and decades to come. In addition, also the
46 chromatographic instrumentation will need to follow these improvements and changes in the
47 performance and format of chromatographic columns [5].

48 Overall, the main factors that determine the separation power and speed of any system are
49 given by the Knox and Saleem equation [6,7], determining the time, t , needed to achieve a given
50 number of theoretical plates, N , under fully optimized kinetic conditions and for a given
51 maximum pressure, ΔP_{\max} , (which can be either column or instrument limited) and mobile phase
52 viscosity, η :

$$53 \quad t = \frac{\eta}{\Delta P_{\max}} \cdot E \cdot N^2 \quad (1)$$

$$54 \quad \text{with } E = h_{\min}^2 \cdot \phi = H_{\min}^2 / K_V \quad (2)$$

55 with H_{\min} and h_{\min} the minimum absolute and reduced plate height, respectively and K_v and ϕ
56 the hydraulic permeability and the flow resistance of the bed, respectively. The importance of
57 the instrumentation (pressure limit, extra column dispersion) and the effect of mobile phase
58 viscosity (LC,GC,SFC) in this equation were discussed earlier [5,8]. The focus of the present
59 contribution will be on the factors grouped in the so-called separation impedance E [6,7],
60 determining the column quality. E is a dimensionless number and is hence independent of the
61 size of the support. It only depends on its shape. In fact, the E -number represents the ability of a
62 given chromatographic support shape to transport the mobile phase through the column with a
63 minimum of dispersion and pressure losses. The smaller E , the smaller these losses, and hence
64 the shorter the time needed to achieve a given N . By far the most important geometrical factor
65 determining the value of E is the external porosity ε , essentially because of its effect on the flow
66 resistance. Computational fluid dynamics simulations on idealized monolithic support structures
67 with varying ε have revealed that, while the domain-size based h does not vary much when ε
68 increases, ϕ can easily decrease with an order of magnitude when the structure becomes more
69 open and, for example, increases from $\varepsilon=40\%$ (=porosity of the packed bed of spheres) to say
70 $\varepsilon=85\%$ [9]. As a consequence, also E and the associated separation time (cf. Eq. 1) can be
71 expected to drop by an order of magnitude. For an increase in $\varepsilon=40\%$ to $\varepsilon=60\%$, ϕ is about 3
72 times smaller, offering 3-fold faster separations than with a packed bed. However, in order to
73 benefit from the shorter analysis times via the reduction of E with increasing porosity, this
74 increase should be accompanied by a significant decrease of the support size. If not, the optimal
75 N value for which Eq. (1) holds increases as well, leading to longer analysis time. Finding ways to
76 simultaneously increase ε and decrease the size of the support elements, while maintaining a
77 good structural homogeneity, mechanical strength and sufficient retention surface, is the key to
78 realize a paradigm shift in the speed and performance of LC columns.

79

80

81

82 2. Packed bed columns

83 The vast majority of chromatographic columns sold nowadays are filled with fully or superficially
84 porous particles [10]. These columns show excellent reproducibility in both performance and
85 selectivity and are available from capillary up to (semi-)preparative scale in a wide range of
86 lengths, packed with different particle sizes and stationary phase chemistries. Whereas $h_{\min}=2$
87 was long considered the practically achievable lower limit for column efficiency for analytical
88 columns (2.1-4.6 mm I.D.), the new generation of superficially porous particles (SPP) allows to
89 achieve h_{\min} values as low as 1.4. Spurred by these developments, attempts have been made to
90 produce fully porous particle batches with a reduced particle size distribution, such that
91 nowadays h_{\min} -values as low as 1.7 can be achieved with fully porous particles [11-14]. It should
92 be noted that in capillary formats, h_{\min} -values down to $h_{\min}=1$ have been demonstrated in
93 research laboratories [15].

94 The fact that particles in a packed bed need to be in contact with each other to obtain a stable
95 and pressure resistant bed, makes that the external porosity ε of a randomly packed bed is
96 always around 36 to 40%. As a consequence, the flow resistance ϕ_0 of packed beds is difficult to
97 alter or optimize and usually lies between 600 and 800 [16,17]. In a first approximation, ϕ_0 can
98 be calculated according to Kozeny-Karman's law (based on the u_0 velocity of an unretained t_0 -
99 marker):

$$100 \quad \phi_0 = 180 \cdot \frac{(1-\varepsilon)^2 \cdot \varepsilon_T}{\varepsilon^3} \quad (3)$$

101 Since ϕ_0 not only depends on ε but also on the total porosity ε_T , a clear reduction of the flow
102 resistance is obtained when switching from fully porous to superficially or non-porous particles
103 (lower total porosity ε_T than fully porous particles). The effect is however rather small and
104 difficult to exploit because a reduction of the porous zone fraction of the particles reduces the
105 sample loadability, causing efficiency loss when a large sample mass is injected [18].

106 The only way to further improve the kinetic performance of packed bed columns would thus be
107 a further reduction in h_{\min} . The latter can be highly effective, given the quadratic variation of E

108 with h_{\min} . As a result of this quadratic dependency, the seemingly modest reduction of h_{\min} by
109 some 20-30% that is typically observed when moving from fully porous to superficially porous
110 corresponds to a very significant two-fold reduction in analysis time. Using typical values for h_{\min}
111 and ϕ_0 for superficially porous ($h_{\min}=1.5, \phi_0=600$) and fully porous ($h_{\min}=2, \phi_0=800$) particle
112 columns, E-numbers are around 1350 and 3200 respectively. These values are in good
113 agreement with experimental results [17,19].

114 Further improvements in packing heterogeneity, reducing the so-called eddy dispersion
115 contribution (A-term), are thus of high interest. As recently discussed by Gritti et al., perfectly
116 ordered packed beds (A-term = 0) are expected to yield h_{\min} values equal to 0.9, 0.7 or 0.5 for
117 fully, superficially and non-porous particle columns respectively, as it is impossible to eliminate
118 longitudinal diffusion and mass transfer resistance contributions [13]. Finding ways to further
119 suppress the eddy-dispersion while sticking to the traditional slurry packing methods seems to
120 be rather difficult (Fig. 1a), if not impossible, given the many efforts already devoted to the
121 problem in the past decades [13]. It seems that radically novel packing methods are needed.
122 One approach, currently under investigation in our group would be the use of additive layer
123 manufacturing, where ordered layers of monodisperse silica particles are assembled layer by
124 layer to form a 3D printed particle column (Fig. 1b). However, this concept is still far from
125 reality. Besides the cost efficiency, one critical aspect is the pressure stability of these beds
126 under the very high operating pressures (up to 1500 bar) nowadays available in commercial
127 UHPLC equipment.

128 An approach to lower the h_{\min} of packed bed columns that appears closer to reality (given the
129 existence of an experimental proof delivered by Wei et al.) is the production of core-shell
130 particles wherein the mesopores are oriented purely radially instead of forming a randomly
131 connected network. Although this seems only a small change, it has such a strong effect on the
132 B-term that it can be expected to lead to a further reduction of 0.5 reduced h-units compared to
133 the conventional core-shell particle performance [20,21].

134

135 **3. Monolithic columns**

136 Instead of stacks of individual particles as in packed bed columns, monolithic columns consist of
137 a continuous porous skeleton with large through-pores (Fig. 2). Already in the early 1950's the
138 potential of this column format was discussed by Nobel Prize Laureates Martin and Synge [22].
139 The in-situ synthesis of monolithic materials has several advantages, such as the absence of frits
140 to retain particles in the column and a facilitated development of miniaturized column formats,
141 such as capillaries (Fig. 2a,2c) and microfluidic chips (Fig. 2e). Also the use of thin monolithic
142 layers to obtain a retentive porous layer for use in open-tubular (Fig. 2d) or pillar-array devices
143 (Figs. 2b) has been demonstrated [23-25]. In principle monolithic stationary phases have the
144 potential to outperform packed columns. Whereas the efficiency of packed columns is related
145 to the particle size while the total porosity and thus flow resistance is fixed, the use of porogenic
146 solvents in the preparation of monolithic materials facilitates the optimization of the globule
147 size/skeleton (almost) independently of ϵ . The advantage that ϵ can be made very large (values
148 up to $\epsilon=86\%$ have been reported for use in liquid chromatography [26]), makes them
149 intrinsically much better suited to obtain small E-values and a correspondingly improved kinetic
150 performance. Silica-based capillary monolithic columns for example have been shown to
151 produce E-values as low as 300 [27]. This quantum leap in E is entirely due to the lower flow
152 resistance of monolithic columns (in turn a direct consequence of their higher external porosity
153 ϵ), because monolithic columns can at best (i.e., when they are produced with similar degrees of
154 eddy-dispersion) be expected to produce about the same (domain size-based) h_{\min} -value as
155 packed bed columns (cf. the small effect of ϵ on h_{\min} in Fig. 10 of [9]).

156 However, an advantageous shape and a concomitantly low E-number is not everything. The
157 absolute size of the support also matters. Here the rule is very simple: instead of creating a large
158 external porosity by increasing the size of the through-pores, the latter should be kept constant
159 (or even made smaller) in order to keep the same mobile to stationary zone diffusion distances.
160 The only way to achieve the required high external porosity then consists of shrinking the size of
161 the structural elements. This however brings about a number of problems which seem so
162 difficult to solve they currently impede the success of monolithic columns. By far the most

163 tenacious problem in this respect is the so-called small domain size limit problem [28,29]. This
164 problem originates from the fact that each monolith synthesis process inevitably displays a local
165 variability on the size and position of the produced solid zone elements, which is at best
166 absolute in size. This implies that the general heterogeneity of the structure will increase when
167 smaller feature sizes are being pursued, putting a fundamental limit on the possible feature size
168 reduction of monolithic columns.

169 **3.1 Polymer monolithic columns**

170 Whereas early forms of (gel-like) polymer monolithic materials collapsed when pressure was
171 applied, rigid polymer-based monolithic materials (Fig. 2c), that are compatible with high-
172 pressure operation, have become available since the 1990s [30,31]. The two most prominent
173 classes of materials are the poly(styrene-co-divinylbenzene) based materials and monolithic
174 entities based on methacrylate-ester-based precursors. Polymerization mixtures are typically
175 prepared from mono- and oligovinyl monomers and an initiator in the presence of an inert
176 diluent, called porogen. The porogen, typically a binary solvent mixture, is selected based on
177 their ability to dissolve the monomers, yielding a homogenous solution. During the course of the
178 polymerization reaction microgel particles are formed, following interparticle reactions via
179 pendant vinyl groups leading to the formation of microgel clusters [32]. Ultimately, a
180 microscopic porous network is formed, and a phase separation occurs. Details of how the
181 reaction conditions affect the size of the microglobules and resulting macropore structure can
182 be found in literature [33,34]. To advance the kinetic performance of monoliths, Vaast et al.
183 described the development of nanostructured high-porosity monolithic supports allowing for
184 sub-min peptide separations [35]. Furthermore, he linked the effects of macropore and
185 microglobule size, and structure homogeneity, to the separation performance measured in
186 gradient elution, both in terms of peak capacity and gradient plate height [35].

187 Polymer-monolithic stationary phases have emerged as an attractive alternative for packed
188 columns in the field of biomolecule separations and their potential has been demonstrated for a
189 wide range of biomolecules [36,37]. In reversed-phase gradient mode, ultra-fast separations (<
190 1 min gradients) of intact proteins have been realized in both large I.D. columns and using

191 capillary column formats [35]. Using a 250 mm long capillary monolithic column and applying a
192 2h gradient, intact proteins, including protein isoforms arising from various amino-acid
193 modifications, were resolved yielding a maximum peak capacity of 650 [38]. Fig. 3 shows the
194 separation of an *E.coli* digest using a 1 m long monolithic column yielding a peak capacity in
195 excess of 1,000 [39]. To further extend the kinetic performance and applicability of monolithic
196 columns, different innovative approaches are being currently explored, such as composite
197 cryopolymers [40] or the incorporation of nanoparticles to extend monoliths with only reversed-
198 phase functionalities to ion-exchange [41]. These nanoparticles might also act as structure
199 directing agents to improve kinetic performance.

200 Although excellent results can be obtained for the separation of larger biomolecules, the plate
201 numbers achieved for small-molecules on polymer-monolithic columns are typically one order
202 of magnitude lower than that obtained on classical packed columns. Whereas the C₁₈ layer on
203 modified silica particles is extremely thin and hence the diffusion distance is short, it has been
204 speculated that small molecules can penetrate into the polymer globules of monolithic
205 materials, and excessive dispersion is a result of 'surface diffusion' [42].

206

207 **3.2 Silica monolithic columns**

208 Silica monoliths (Fig. 2a) are produced via a sol-gel process wherein alkoxy silanes are hydrolyzed
209 and then polycondensed in the presence of a water-soluble porogen [43, 44]. Siloxane
210 oligomers formed during successive condensation reactions link together to form a gel network.
211 Spinodal decomposition occurs and phase separation takes place between the silica-rich and
212 solvent-rich phase, forming the future silica skeletons and through-pores, respectively. Similar
213 to polymer monoliths, the phase separation and the pore size of the gel are controlled by
214 varying the concentration of the porogen. The stiffness and strength of the gel are increased by
215 aging in a siloxane solution and mesopores are formed by adding ammonium to the aging
216 solution. Finally, the gel is dried and clad with polyether ether ketone (PEEK) to obtain a silica
217 monolith suitable for chromatographic purposes. This column housing however limits the
218 maximum operating pressure in analytical scale columns to HPLC-like operating pressures

219 (generally below 400 bar), while it has recently been shown the silica monolithic skeleton itself
220 can withstand pressures up to at least 800 bar [45].

221 For analytical scale monoliths (2.1-4.6 mm I.D.), through-pores sizes are typically $d_{tp}= 1-2 \mu\text{m}$
222 and high external porosities ($\varepsilon > 60\%$) are obtained. Due to their intrinsic high permeability,
223 silica monoliths can be operated at high linear velocities, or in long (coupled) columns, resulting
224 in extremely high efficiencies [46]. The small size of the silica skeletons (typically $d_{skel}= 1-2 \mu\text{m}$)
225 results in efficiencies comparable to those obtained in columns packed with $5 \mu\text{m}$ particles,
226 especially when operated at high flow rates [47]. However, due to the poor radial homogeneity,
227 which can be related to their fabrication process, concomitant high eddy dispersion and their
228 limited pressure resistance, silica monoliths [48] have not been able to compete with the
229 particle-packed columns (sub- $2 \mu\text{m}$ or sub- $3 \mu\text{m}$ core-shell) that were developed around the
230 same time [49-51]. To improve their performance, efforts have been made to improve the radial
231 homogeneity while at the same time reducing their feature sizes by adjusting the preparation
232 process (e.g. concentration and porogen type). This has resulted in the introduction of the so-
233 called second generation of silica monoliths [27,52,53]. Due to their improved radial
234 homogeneity and reduced skeleton ($d_{skel}<1 \mu\text{m}$) and through-pore sizes ($d_{tp}=1.1-1.2 \mu\text{m}$), H_{min}
235 values are much lower compared to the first generation, and comparable to what can be
236 obtained in $3-3.5 \mu\text{m}$ particle packed columns [54]. The downside of these reduced feature sizes
237 is that the permeability of the monolithic column decreases accordingly, from $K_{v0}=4.7 \cdot 10^{-14} \text{ m}^2$
238 and $K_{v0}=4.0 \cdot 10^{-13} \text{ m}^2$ for the first generation [26,27,55] to significantly smaller values $K_{v0}=2.0 \cdot 10^{-$
239 $^{14} \text{ m}^2$ for the second generation [55]. According to Deridder et al., the permeability of a silica
240 monolith is directly related with the square of its skeleton size, while a more complex relation
241 between permeability and external porosity exists, depending on the geometry of the monolith
242 [56]. Considering that external porosity values measured for first and second generation
243 monoliths are largely the same, the decreased permeability of the second generation monoliths
244 must therefore mainly be attributed to the reduced skeleton and through pore sizes [55].
245 Nevertheless, the lower permeability of the second generation monoliths is still well above
246 those measured for sub- $3 \mu\text{m}$ particle columns.

247 Comparing silica monoliths with packed bed columns, similar E-values (at the lower end of the
248 range) as for fully porous particles columns are found, with $E=2200-4600$ and $2200-3400$ for the
249 first and second generation, respectively [55]. To compare the separation power for a given
250 separation problem, the kinetic plot method is a useful alternative to the impedance, as it
251 represents the maximum plate count obtainable in a certain analysis time. Fig. 4 compares the
252 kinetic performance for first and second generation monoliths with $\Delta P_{\max}=200$ bar. It is clear
253 that second generation monoliths perform better (1.5-2.5x faster for a certain N) than the first
254 generation monoliths for $N<50.000$. For more challenging separations ($N > 50.000$), the first
255 generation monoliths perform better as their large permeability allows them to be used in
256 longer columns without compromising the separation speed [55]. Comparing their performance
257 with a $2.7 \mu\text{m}$ superficially porous particle packed column, operated at its own $\Delta P_{\max}=600$ bar,
258 shows that the SPP-based column outperforms both generations of monoliths over the entire
259 range of practically relevant plate counts. A similar conclusion can be drawn when comparing
260 silica monoliths with sub- $2 \mu\text{m}$ particle columns with $\Delta P_{\max}=1000-1500$ bar [57]. Further
261 improvements in the structural size and homogeneity of the silica monoliths by improving their
262 production process, together with the development of higher pressure-resistant material to clad
263 the monolithic columns, are required before the monolithic columns can become competitive
264 with the current state-of-the-art in particle packed column technology.

265 In capillary formats, the radial variation in external porosity (and hence in flow resistance)
266 caused by the inevitable post-synthesis shrinking process is much smaller than in analytical bore
267 columns. Producing silica monoliths with a domain size of about $2 \mu\text{m}$ in $100 \mu\text{m}$ columns, H_{\min} -
268 values as low as $4.1-4.4 \mu\text{m}$ have been demonstrated [58]. This is however still around two
269 times larger than the lowest H_{\min} ever reported for packed bed columns ($H_{\min}=2 \mu\text{m}$ when using
270 $1.3 \mu\text{m}$ core-shell particles) [59]. This shows a further significant decrease of the domain size of
271 silica monolithic columns is still needed.

272

273

274 **4. Microfabricated columns**

275 Pillar array columns were introduced in 1998 by Fred Regnier and co-workers as an ordered
276 alternative to disordered chromatographic packings [60,61]. Because the packing was originally
277 intended for capillary electro-chromatography separations requiring non-conducting substrates,
278 the first experiments were carried out using columns produced in fused silica and PDMS.
279 Because these substrates do not easily allow fabrication of pillars with sidewall slopes close to
280 90°, the substrates had to be replaced by silicon by other researchers in follow-up studies
281 before the predicted absence of eddy dispersion due to the perfectly ordered structures was
282 indeed reflected in the measured van Deemter curves. The first reversed-phase separations on
283 silicon micro-pillar arrays were reported in 2007 [62], showing plate heights as low as 4 µm for
284 retained components in a non-porous pillar bed. These initial results were obtained by
285 measuring the band broadening in the center of the beds, i.e., by excluding the sidewall region
286 where the flow resistance of the bed was different from that in the rest of the bed. Using
287 computational fluid dynamics simulations this problem could be solved and appropriate designs
288 for the side-wall region were proposed [63]. One particularly useful solution were radially
289 elongated pillars (REP, Fig. 5g) having a lateral to axial aspect ratio larger than 10 [64-66]. The
290 use of (at least a number of rows of) such REP structures in the flow distributors (Fig. 5f) at the
291 inlet and outlet sections of the bed also proved to be essential to interface the columns with the
292 outer world [67]. These distributors were also key to produce sufficiently long columns on the
293 (relatively limited) surface of a silicon wafer, connecting different channel tracks using low
294 dispersion turns (Fig. 5b). An interesting alternative approach was implemented by Isokawa *et*
295 *al.* [68], who designed a dedicated curve and pillar bed with varying density to minimize
296 dispersion, while at the same time increasing permeability in the turn zone.

297 In 2017, the first generation of a pillar array columns was introduced, consisting of a 2 m long
298 pillar bed with 5 µm diameter pillars, spaced 2.5 µm from each other (width 315 µm, depth 18
299 µm, porous layer of 200 nm) (Figs. 5a-5c). This column has the permeability of a 10 µm packed
300 bed (i.e. $2.7 \cdot 10^{-13} \text{ m}^2$), while it produces plate heights comparable with 3-4 µm porous spherical
301 beads (i.e. 5-7 µm) [67]. The corresponding E-number is on the order of 50 to 100, i.e., more

302 than an order of magnitude less than in packed bed columns. The pressure tolerance of more
303 than 400 bar would even allow to construct columns of more than 10 m, producing more than
304 1.5 M plates under retained conditions (small molecules like phenones) in about half a day
305 (12h). A lot of efforts were put in the conformal integration of porous layers into the chips, in
306 order to increase the specific surface of the non-porous pillars. Two methods that additionally
307 allow tuning of the pore geometry have been developed to this end and have been applied for
308 2.5 μm pillar spacings [24,69]. With electrochemical anodization (silicon) pores are grown inside
309 the silicon pillar (Fig. 5d-5e), leaving the contour of the pillar unaltered [69], whereas with the
310 sol-gel deposition technique a porous glass layer is grown on the pillar (Fig. 2b), thereby
311 increasing the size of the pillar [24]. For some applications, such as ion-pair reversed-phase
312 chromatography of nucleotides [70] or hydrodynamic chromatography [71], it is actually
313 preferred that the pillars are non-porous (Fig. 5b) [72]. In this case, the plate count roughly
314 doubles [73]. These columns have an equivalent cylindrical diameter of around 80 μm , which is
315 a typical targeted diameter to achieve optimal flow rates for ESI-MS detection. These column
316 features make the 1st generation of commercial pillar array columns extremely suited to achieve
317 peak capacities > 1000 in the nano-LC flow rate range.

318 Since this 1st generation still uses relatively large pillars (5 μm) great improvements in both
319 speed and efficiency can be expected when new generations will be produced with pillar
320 diameter and spacing of the same order as the current sub-2 μm particles used in packed bed
321 columns. Technologically this is feasible, given the Bosch etching technology has the potential to
322 achieve even sub-micron resolution [63]. Sidewall effects might appear again, but this could be
323 countered by using REP structures that are insensitive to this effect [65].

324

325 **5. 3D-printed columns**

326 In the last decade, almost every field in scientific research has started to use 3D printing or
327 additive manufacturing. Compared to traditional manufacturing technologies, 3D printing offers
328 the possibility to use the full three-dimensional fabrication potential to freely tune and fabricate
329 any favorable geometry. For applications in chromatography, 3D printing especially holds the

330 promise of providing a way to produce perfectly ordered structures, thus allowing to eliminate
331 any eddy-dispersion contributions. Similar to micro-fabricated columns, the external porosity
332 and thus the flow resistance can freely be tuned within the range where mechanically stable
333 structures can be printed. Combining 3D printing with computational fluid dynamics simulations
334 can lead to the design of “fully optimized” stationary phases, having limited diffusion and a low
335 resistance to mass transfer. Another asset of 3D printing is the fast manufacturing of
336 prototypes, making rapid iterative device development possible. The use of additive
337 manufacturing should not be limited to stationary phases, as complete chromatographic
338 columns (containing stationary phase, column wall, flow inlet and frits) can be fabricated
339 simultaneously. Combining 3D printed columns with printed valves, micropumps, connectors
340 and other microfluidic parts [74], one can even dream of producing a complete 3D printed
341 chromatographic system on a chip.

342 The emerging possibilities of 3D printing have already reached the field of HPLC as could be
343 seen at the latest HPLC conferences. In 2016, Brett Paul and co-workers showed the possibilities
344 to fabricate 3D metal printed chromatographic columns, which were functionalized in-situ with
345 a thermally polymerized monolith [75]. Instead of using 3D printing to obtain a scaffold on
346 which a stationary phase is deposited, the whole column, including walls, can be printed in one
347 step, as shown by Fee et al.. They exploited the full 3D-potential of additive manufacturing,
348 printing both octahedral beads in a simple cubic configuration (apothems of $113.6 \pm 1.9 \mu\text{m}$), and
349 monolith hexagonal channels, both in parallel and herringbone arrangements (apothems of
350 $148.2 \pm 2.0 \mu\text{m}$) [76]. In 2017, his group printed a broad range of particle shapes including
351 tetrahedral, octahedral, stellar octangular, triangular bipyramid and truncated icosahedra
352 particles, in different geometric arrangements as simple cubic, body-centered cubic and face-
353 centered cubic [77]. However, for 3D printing to become the new standard of column
354 manufacturing, some significant disadvantages need to be overcome. The most important
355 hurdle is the resolution or minimal printed feature size, being around 25-100 μm for the most
356 widespread technologies, such as extrusion based printing (Fused Deposition Modeling),
357 stereolithography (Stereolithographic Apparatus, Digital Light Processing), and powder-based
358 printing (Selective Laser Melting, Selective Laser Sintering, inkjet-based printers) [74]. State-of-

359 the art packed bed column technologies with e.g. sub-2 μm particles have feature sizes (flow-
360 through pores) in the order of 500nm and below. Currently, only one additive manufacturing
361 technology exists which offers competitive resolutions, namely two-photon polymerization
362 printing (2PP). Nevertheless, other 3D print technologies with lower resolutions can be applied
363 for the manufacturing of preparative columns, not being so demanding towards micron-scale
364 feature sizes. A 2PP printer operates by emitting near-IR femtosecond pulses of photons into a
365 photo-polymerizable resin. Two photons need to be simultaneously absorbed to initiate radical
366 polymerization, a process so rare that it only takes place in the focal spot size of the laser,
367 leading to extremely small voxel (volume-pixel) sizes. With minimal feature sizes smaller than
368 50nm [78], stationary phases with characteristic distances of 1 μm size can thus be
369 manufactured with high accuracy (see Figs. 2f and 6). A drawback of 2PP however is the trade-
370 off between high resolution and printing time/volumes. Whereas new structures can be quickly
371 prototyped in manifold with all other 3D print technologies, the speed of (mass-)production
372 with 2PP still has a long way to go. The manufacturing of a structure as shown in Fig. 2f ($\epsilon=80\%$,
373 edges of 1 μm , through pores of 1.5 μm) for a 1cm long column with a width of 100 μm and 10 μm
374 deep (equivalent to a 40 μm ID capillary) takes almost 1 day to print. It needs to be mentioned
375 that printing time is highly influenced by the spatial dimensions, structure, material choice, laser
376 objective, writing direction and many other parameters. In addition, 2PP is much more
377 expensive than the more widespread technologies.

378 Another important hurdle is the material choice, as the final obtained structure needs to be
379 temperature and solvent resistant (e.g. no swelling). In addition, these materials will have to be
380 functionalized to obtain an appropriate retentive surface chemistry (e.g. C₈, C₁₈, phenyl,
381 amide...), to such an extent that no undesired interactions are possible with the starting
382 material. As typical 2PP printed material have no inherent porosity, either the printed structure
383 will need to be modified to create mesopores or a porous layer will need to be deposited on the
384 surface of the printed structure to obtain sufficiently high retention surface.

385

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387

388 **6. Conclusions:**

389 Despite the many research efforts on novel column technologies, packed columns still are the
390 first choice in liquid chromatography. Monolithic columns are rather employed in niche
391 applications or for their flexible applicability and possible in-situ generation in complex
392 geometries. Further improvements in the production process of the silica-gel monoliths should
393 aim at reducing the domain size further while maintaining or further improving the
394 homogeneity before they can ever become competitive with the current state-of-the-art
395 particle packed columns. The development of a suitable ultra-high pressure column housing is
396 another issue. Polymeric monolithic columns show great potential for the separation of large
397 (bio)molecules but have inherent disadvantages for small molecule separations. The first
398 generation of microfabricated pillar array shows promising results for separations that require
399 high resolving power, but reduced feature sizes are required to further enhance separation
400 speed and efficiency. 3D printing shows a tremendous intrinsic potential for the fabrication of
401 chromatographic columns, but the current printing hardware either does not allow to obtain
402 the desired resolution (sub- μm) or requires too long printing time, limiting the technology at
403 this moment to either preparative scale separations or the development of chip-scale devices.
404 These novel developments clearly show that we have not yet reached the end of column
405 development and further improvements can be expected in upcoming decades.

406

407

408 **Figure Captions**

409 Fig. 1: (a) SEM image of monodisperse particles in a random sphere packing showing the
410 inherent packing heterogeneities; (b) artist's impression of packed bed column made using
411 additive layer manufacturing, with monodisperse particles assembled layer by layer to form a
412 3D printed particle column.

413 Fig. 2: SEM images of (a) silica monolithic column, (b) silica monolithic layer deposited on REP
414 column [24], (c) polymer monolithic column, (d) silica monolithic layer deposited on capillary
415 column for use in open tubular LC [23], (e) silica monolithic column synthesized in pillar array
416 column [25], (f) 3D printed monolithic column. Figures adapted from refs., with permission.

417 Fig. 3: High-resolution LC-MS/MS analysis of a tryptic digest of E. coli obtained on a 1 m long
418 monolithic column. Adapted from ref. [39], with permission.

419 Figure 4: Kinetic plots of analysis time (t_R) versus plate count (N) for benzophenone and for first
420 generation monoliths (\diamond and Δ) and second generation monoliths (\blacksquare \blacklozenge \bullet \blacktriangle) evaluated at 200
421 bar. Open, blue symbols refer to first generation monoliths, closed, black symbols to second
422 generation monoliths. The red curves (\times) are obtained for a core-shell column (2.0x100mm,
423 $d_p=2.7 \mu\text{m}$) operated at a maximum pressure of 600 bar. The mobile phase was adapted on all
424 columns to obtain $k=8.7$ for benzophenone. Adapted from [55], with permission.

425 Fig. 5: (a) Silicon waver featuring several pillar array columns, with zoom in on (b) turn
426 structures at the end of each channel to increase total column length, (c) cylindrical pillars
427 making up the chromatographic bed, (d) anodized pillars to increasing retentive surface [67], (e)
428 detail of porous shell [67], (f) channel inlet flow distributor; (g) alternative bed structure with
429 radially elongated pillars [65]. Figures adapted from refs., with permission.

430 Fig. 6: Example of a 3D printed tetrahedron skeleton model type bed (zoom: see Fig. 2f).

431

Figure 1:

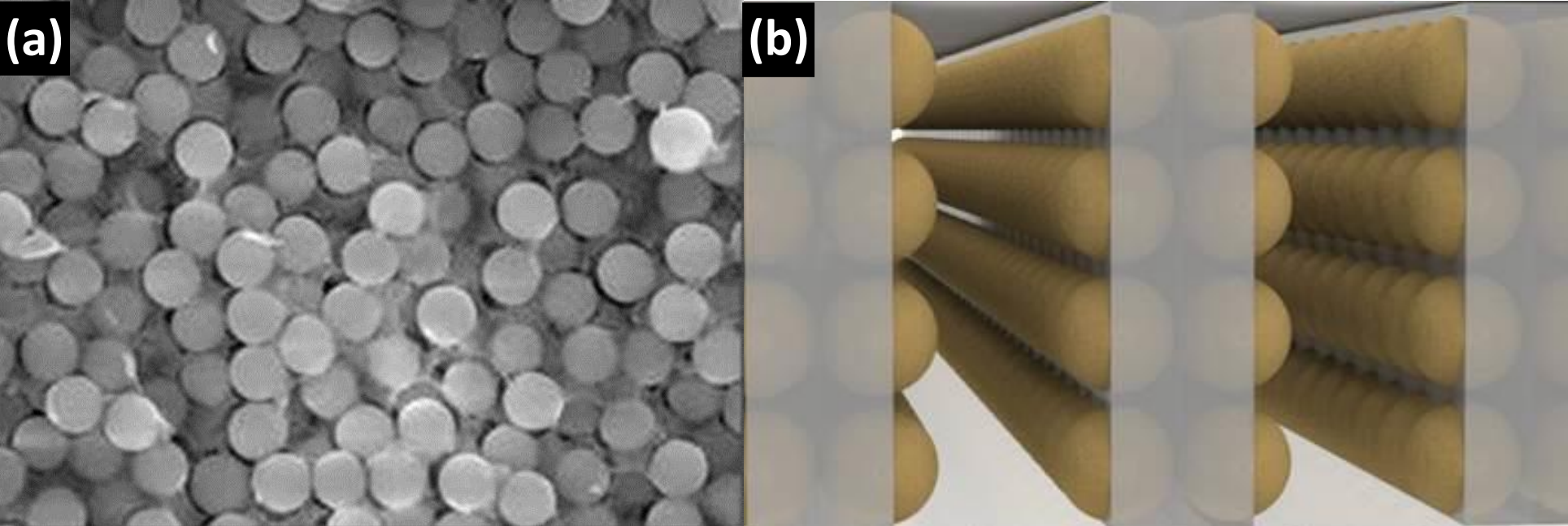


Figure 2:

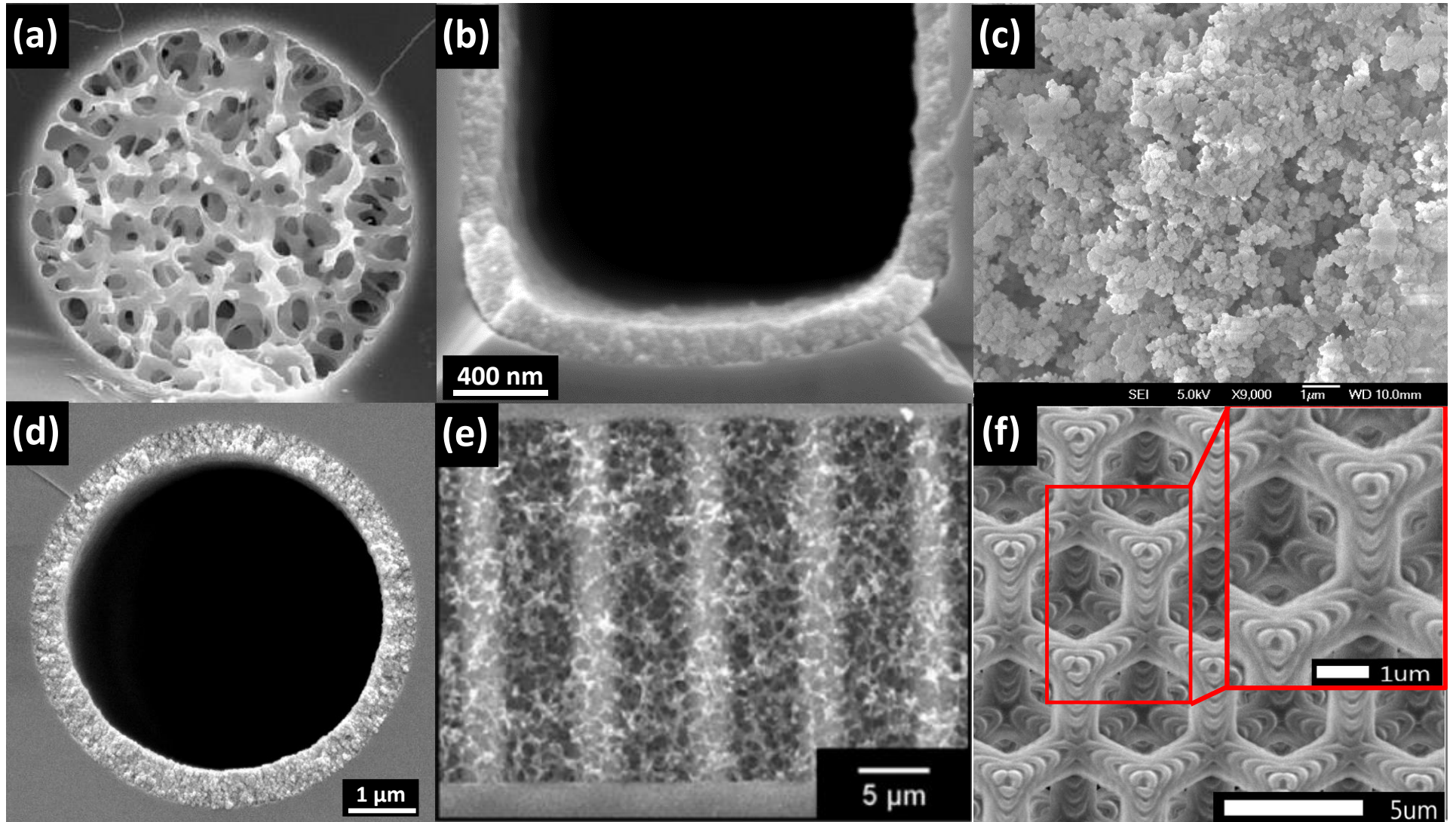


Figure 3:

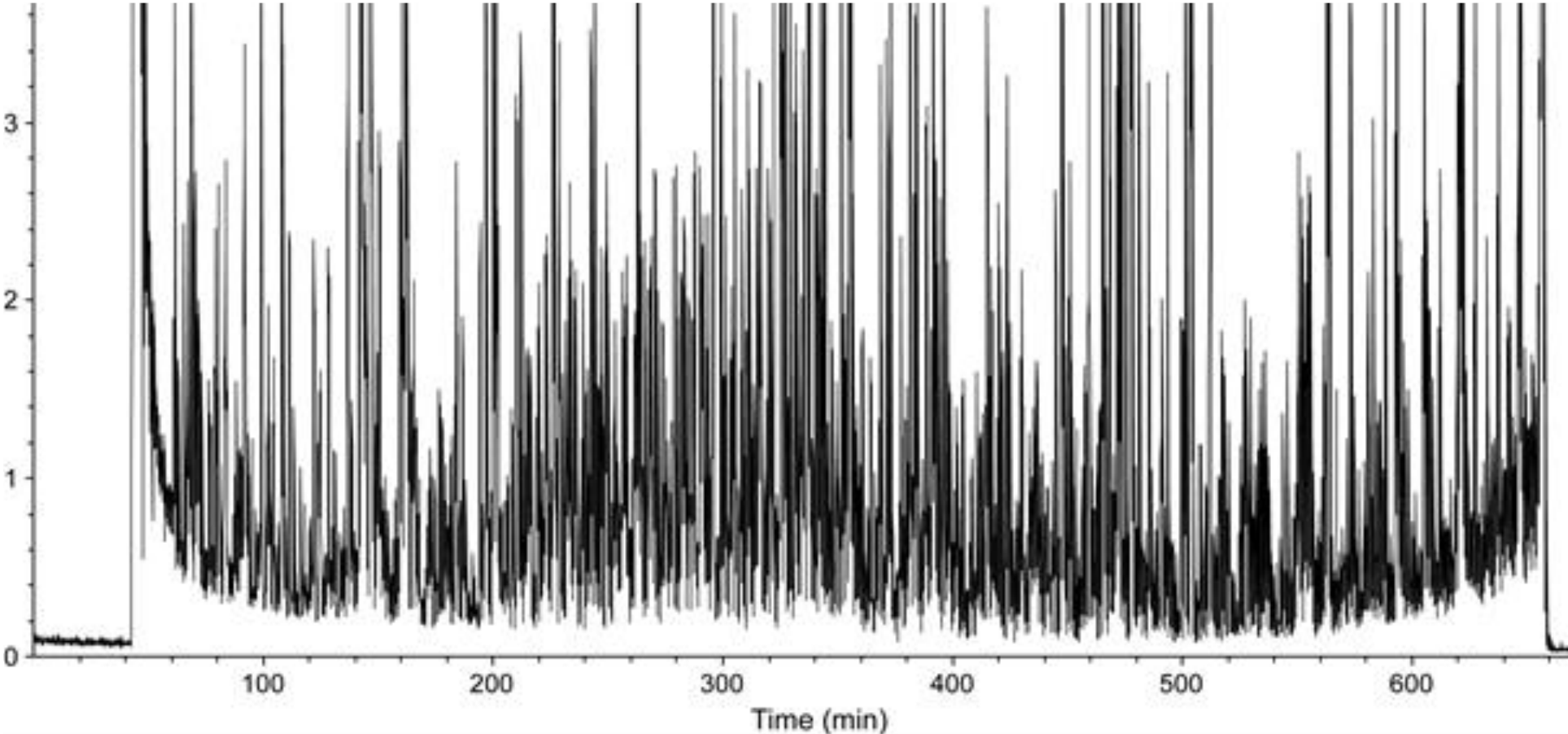


Figure 4:

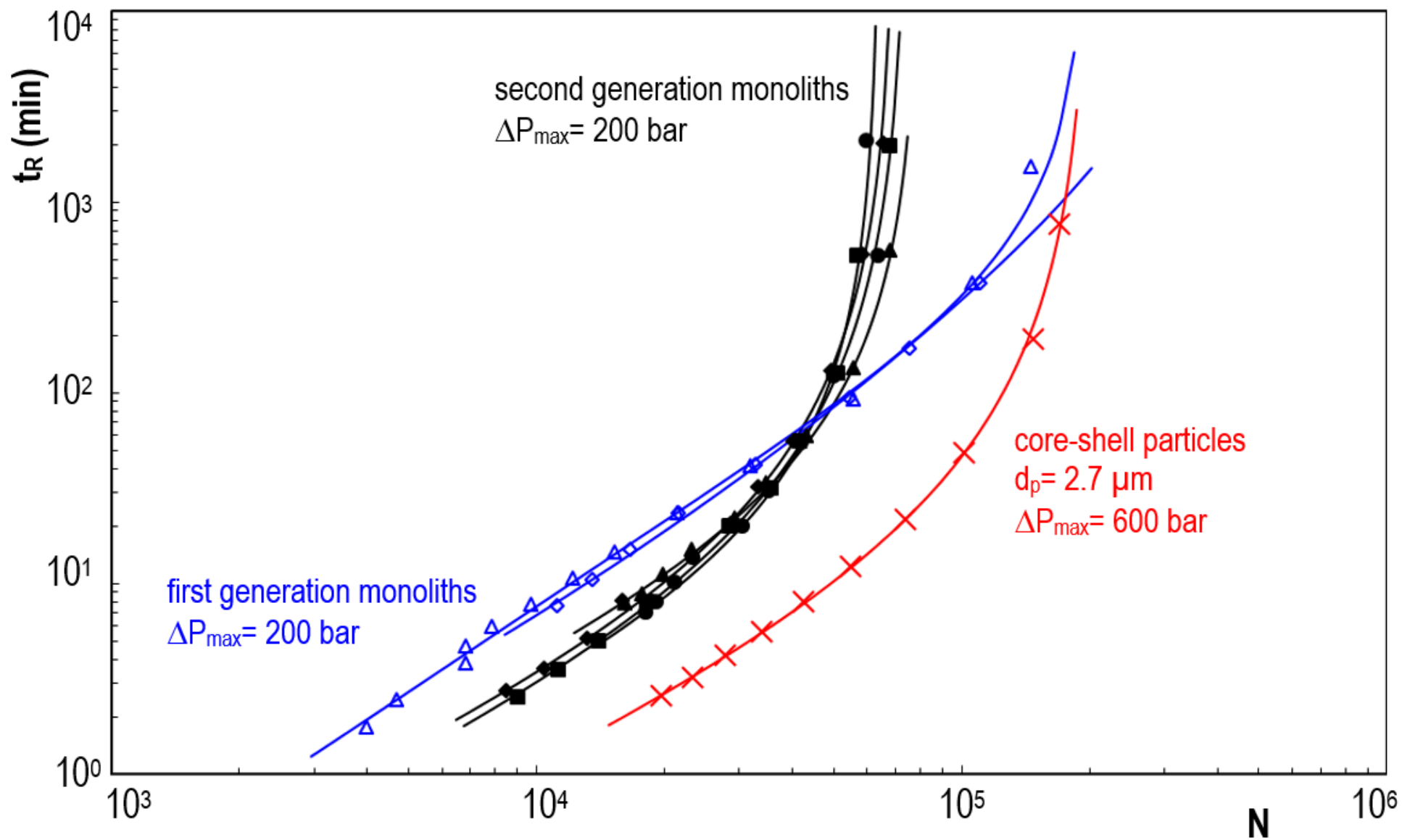


Figure 5:

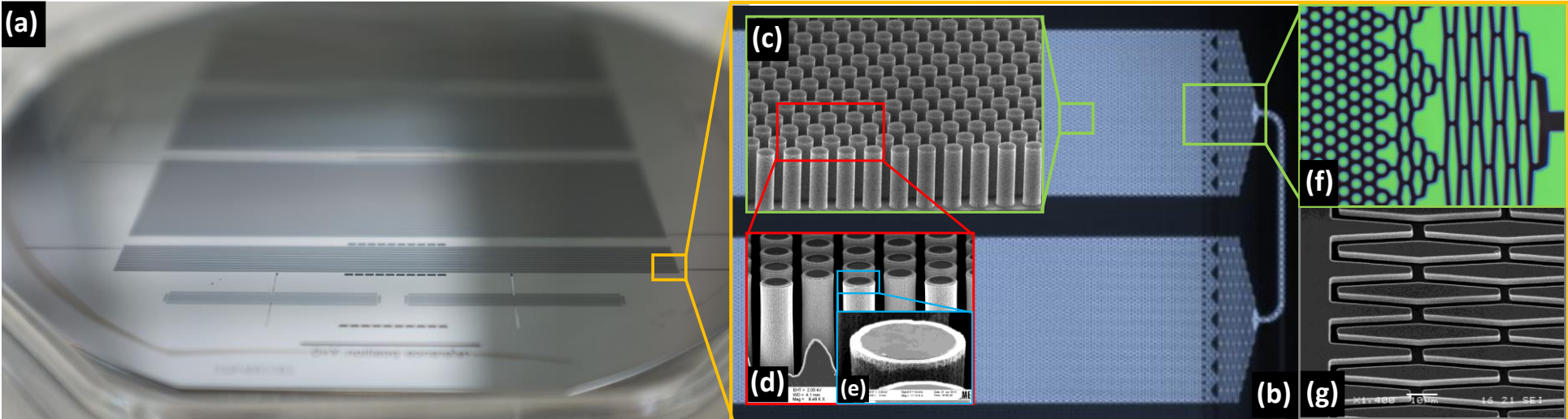
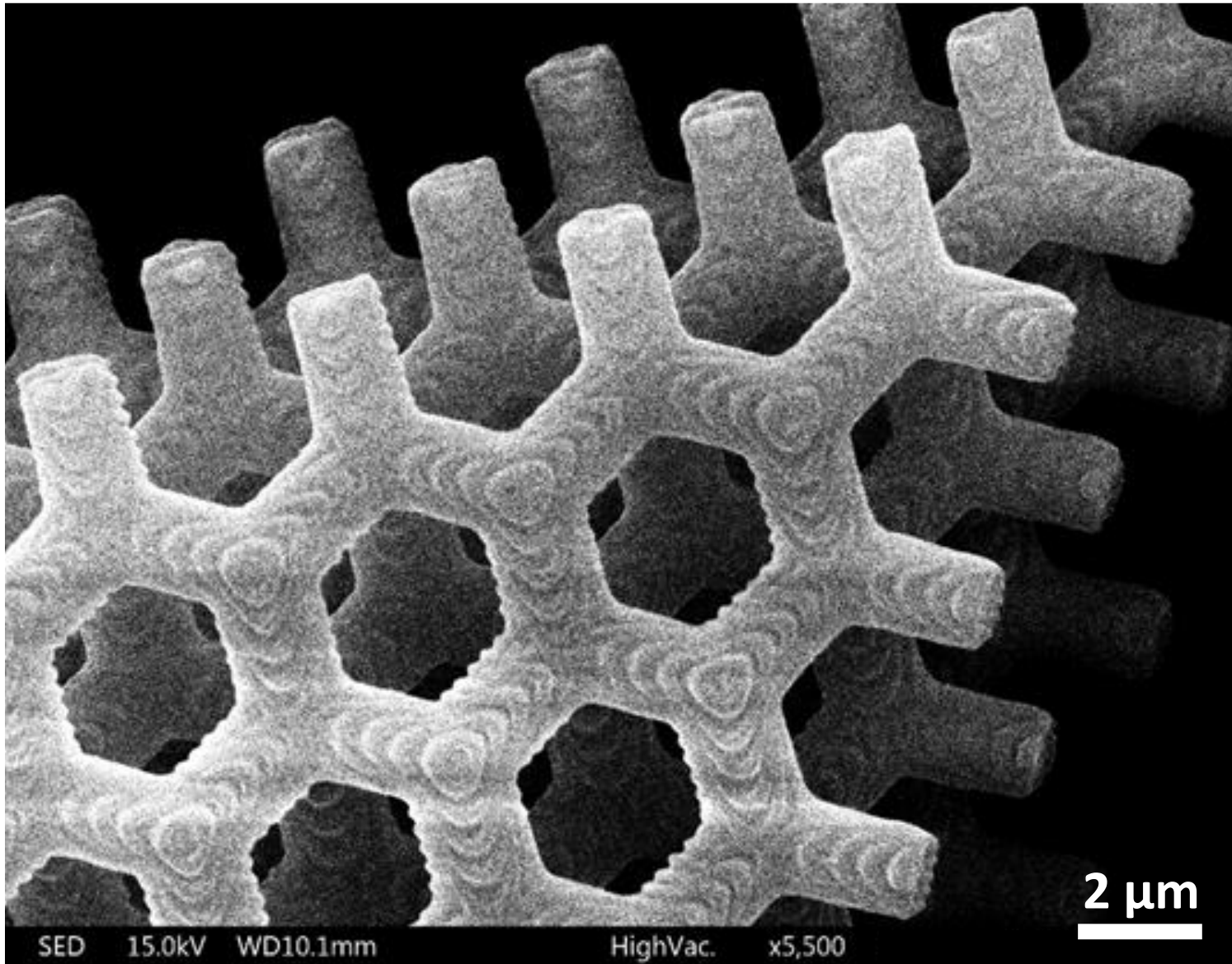


Figure 6:



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